REPORTS

References and Notes

- 1. J. A. Engelman, J. Luo, L. C. Cantley, *Nat. Rev. Genet.* 7, 606 (2006).
- 2. P. V. Schu et al., Science 260, 88 (1993).
- P. Burda, S. M. Padilla, S. Sarkar, S. D. Emr, J. Cell Sci. 115, 3889 (2002).
- 4. O. V. Vieira et al., J. Cell Biol. 155, 19 (2001).
- 5. I. Vergne *et al., Proc. Natl. Acad. Sci. U.S.A.* **102**, 4033 (2005).
- A. Simonsen, S. A. Tooze, J. Cell Biol. 186, 773 (2009).
- M. P. Byfield, J. T. Murray, J. M. Backer, J. Biol. Chem. 280, 33076 (2005).
- T. Nobukuni *et al.*, Proc. Natl. Acad. Sci. U.S.A. 102, 14238 (2005).
- J. E. Slessareva, S. M. Routt, B. Temple, V. A. Bankaitis, H. G. Dohlman, *Cell* **126**, 191 (2006).
- 10. J. M. Backer, Biochem. J. 410, 1 (2008).
- 11. C. Panaretou, J. Domin, S. Cockcroft, M. D. Waterfield,
- J. Biol. Chem. **272**, 2477 (1997). 12. Y. Yan, R. J. Flinn, H. Wu, R. S. Schnur, J. M. Backer, *Biochem. J.* **417**, 747 (2009).
- 13. H. W. Shin *et al.*, *J. Cell Biol.* **170**, 607 (2005).
- 14. J. T. Murray, J. M. Backer, *Methods Enzymol.* **403**, 789 (2005)

- K. Obara, T. Sekito, Y. Ohsumi, *Mol. Biol. Cell* **17**, 1527 (2006).
- 16. E. L. Axe et al., J. Cell Biol. 182, 685 (2008).
- 17. G. M. Fimia et al., Nature 447, 1121 (2007).
- A. Kihara, T. Noda, N. Ishihara, Y. Ohsumi, J. Cell Biol. 152, 519 (2001).
- E. Itakura, C. Kishi, K. Inoue, N. Mizushima, *Mol. Biol. Cell* 19, 5360 (2008).
- 20. Q. Sun et al., Proc. Natl. Acad. Sci. U.S.A. 105, 19211 (2008).
- 21. C. Liang et al., Nat. Cell Biol. 8, 688 (2006).
- 22. Y. Takahashi et al., Nat. Cell Biol. 9, 1142 (2007).
- 23. C. Liang et al., Nat. Cell Biol. 10, 776 (2008).
- 24. Materials and methods are available as supporting material on *Science* Online.
- 25. T. Bondeva et al., Science 282, 293 (1998).
- E. H. Walker, O. Perisic, C. Ried, L. Stephens, R. L. Williams, *Nature* **402**, 313 (1999).
- C.-H. Huang et al., Science **318**, 1744 (2007).
 Y. V. Budovskaya, H. Hama, D. B. DeWald, P. K. Herman,
- J. Biol. Chem. 277, 287 (2002). 29. A. Berndt *et al., Nat. Chem. Biol.* 6, 117 (2010).
- Single-letter abbreviations for the amino acid residues are as follows: A, Ala; C, Cys; D, Asp; E, Glu; F, Phe; G, Gly; H, His; I, Ile; K, Lys; L, Leu; M, Met; N, Asn; P, Pro; Q, Gln; R, Arg; S, Ser; T, Thr; V, Val; W, Trp; and Y, Tyr.
- 31. We thank European Synchrotron Radiation Facility beamline scientists for ID14-4, ID23-1, and ID29. We are grateful to Y. Ohashi and A. Gillingham for help with yeast experiments, C. Sachse for electron microscopy, M. Allen for help with x-ray data collection, A. Berndt for advice and discussions, S. Munro for critically reading the manuscript, B. González for a clone of HsVps34, and G. Ducker for help in optimizing Vps34 inhibitor assays. B.T.H. is supported by the Mount Zion Health Fund, B.T. is supported by the Achievement Rewards for College Scientists Fellowship at UCSF, and K.M.S. thanks the Waxman Foundation for support. The coordinates have been deposited in the Protein Data Bank, with the following identification numbers (PDB IDs): 2X6H (apo), 2X6J (PIK-90), 2X6J

Supporting Online Material

www.sciencemag.org/cgi/content/full/327/5973/1638/DC1 Materials and Methods Figs. S1 to S11 Table S1 References

9 November 2009; accepted 26 February 2010 10.1126/science.1184429

Evolutionary Trade-Offs in Plants Mediate the Strength of Trophic Cascades

Kailen A. Mooney,¹* Rayko Halitschke,² Andre Kessler,² Anurag A. Agrawal^{2,3}

Predators determine herbivore and plant biomass via so-called trophic cascades, and the strength of such effects is influenced by ecosystem productivity. To determine whether evolutionary trade-offs among plant traits influence patterns of trophic control, we manipulated predators and soil fertility and measured impacts of a major herbivore (the aphid *Aphis nerii*) on 16 milkweed species (*Asclepias* spp.) in a phylogenetic field experiment. Herbivore density was determined by variation in predation and trade-offs between herbivore resistance and plant growth strategy. Neither herbivore density nor predator effects on herbivores predicted the cascading effects of predators on plant biomass. Instead, cascade strength was strongly and positively associated with milkweed response to soil fertility. Accordingly, contemporary patterns of trophic control are driven by evolutionary convergent trade-offs faced by plants.

rophic cascades-the indirect positive effect of predators on plant biomass through herbivore suppression-are the best examples of the importance of indirect interactions as determinants of community structure and ecosystem function. For this reason, there has been great interest in elucidating the sources of variation in trophic cascade strength both within (1-3) and among ecosystems (4). Much of the research aimed at explaining variation in trophic cascade strength has focused on factors mediating the top-down effects of predators on herbivores, including the influences of intraguild predation (5), synergistic and antagonistic effects of multiple predators (6), trophic subsidies to predators (7), and the nonconsumptive effects of predators on herbivores

(8). At the same time, it is also recognized that plant stoichiometry (9), antiherbivore defense traits (10-12), and primary productivity (13, 14) can mediate trophic cascade strength from the bottom up. Consequently, a consensus is emerging that multiple, complementary top-down and bottom-up processes determine trophic cascade strength.

Although it is recognized that plant traits can influence interactions with herbivores and herbivore-predator interactions (15, 16), there has been little consideration of how plant growth and defense strategies might result in predictable patterns of trophic cascade strength. There is wide acceptance that plant species evolve in response to fundamental trade-offs that should influence the effects of predators and productivity upon herbivore and plant biomass. For example, plant defense theory predicts that fast-growing species should have relatively low herbivore resistance as compared with slow-growing species (17, 18). Plant resistance to herbivores may in turn influence the indirect effects of predators on plants by altering herbivore susceptibility to predators (19, 20). Similarly, plant growth strategies influence tolerance to herbivory (21, 22), again showing potential to alter the strength of trophic cascades. Although trophic cascades are rightly considered community-level phenomena (23), an understanding of how plant traits influence such dynamics requires first documenting the influence of plant traits on component, species-level cascades.

We conducted a field experiment in which we grew 16 species of milkweeds (Asclepias spp., Apocynaceae) (Fig. 1), factorially manipulated predator access and soil fertility, and monitored plant biomass and populations of the potent herbivore Aphis nerii (Aphididae, Hemiptera), a specialist on the Apocynaceae that occurs naturally on the studied milkweeds (24). It has previously been shown that milkweed species influence this aphids' population dynamics and interactions with parasitoids (16, 25). We tested whether there are indirect consequences of such variation in trophic dynamics for plant growth as well as whether trade-offs between milkweed growth strategy and herbivore resistance predictably influence the top-down effects of predators. Because all plants were grown in a single environment, any variation in the effects of predators and growth strategy can be attributed to plant species traits. By interpreting these patterns of interspecific variation in trophic structure from a phylogenetic perspective, we link the outcome of fundamental evolutionary trade-offs to contemporary community dynamics.

We first tested for variation among milkweed species in the effects of predators and soil fertility on both plant biomass and herbivore abundance using general linear models. Where species varied in such responses, we then quantified effect sizes for individual species [log response ratios (26)] in order to examine the relationships among species using phylogenetically independent contrasts (27).

¹Department of Ecology and Evolutionary Biology, University of California, Irvine, CA 92697–2525 USA. ²Department of Ecology and Evolutionary Biology, Cornell University, Ithaca, NY 14853– 2701 USA. ³Cornell Center for a Sustainable Future, Ithaca, NY 14853, USA.

^{*}To whom correspondence should be addressed. E-mail: mooneyk@uci.edu

The 16 milkweed species varied in herbivore resistance (quantified here as $-1 \times$ aphid density in the absence of predators) (fig. S1 and table S1) and the top-down effects of predators on herbivore density and plant biomass (Fig. 1 and table S1). This variation in top-down control among species is equivalent in magnitude to that previously observed among ecosystem types (*4*). That such variation occurs among closely related species in a single abiotic environment, with the same herbivore species and with a common guild of predators, underscores the powerful influence of plant traits upon trophic structure.

Milkweed species also differed strongly with respect to two aspects of the plants' growth strategy, growth rate [which is defined as mean species biomass at the conclusion of the experiment (fig. S1 and table S1)], and the growth response to increased soil fertility (Fig. 1 and table S1). The strength of soil fertility effects on plant biomass was stronger when predators were absent than when present (fig. S2 and table S1), but these dynamics were consistent among milkweed species (table S1). Despite the speciesspecific effects of soil fertility on milkweed growth, the indirect positive effect of soil fertility on aphid density was indistinguishable among milkweed species (Fig. 1 and table S1). Accordingly, there was asymmetry in how milkweed species influenced top-down and bottom-up trophic dynamics: Predator effects on both herbivore density and plant biomass were speciesspecific, whereas species variation in soil fertility

Fig. 1. Effect sizes for influence of predators and fertilization on herbivore density (calculated per gram of plant dry biomass) and final plant biomass. Black bars for A. nivea and A. candida indicate that soil fertility was not manipulated for these species. Effects are natural log response ratios (LRRs) with 95% confidence limits. Predator effects are calculated across both levels of soil fertility, soil fertility effects are calculated across both levels of predation, and all effects are based on manipulation and control sample sizes of n = 10 plants each. LRRs of 1.0, 2.0, and 3.0 correspond to changes of 2.7fold, 7.4-fold, and 20-fold, respectively. Predator effects on both herbivore density and plant biomass differ significantly among species, whereas fertilization effects differ for plant biomass but not for herbivore density (table S1). To the left of the effect sizes, the phylogenetic relationship of the studied milkweeds is presented (29).

effects were limited to the direct influence on plant biomass.

Having shown milkweed species differences in growth strategies, resistance, and the top-down effects of predators, we examined the relationships among these species traits while controlling for phylogenetic history (27). Variation in herbivore density among milkweed species was determined by means of a combination of top-down and bottom-up processes. Although theory and data predict that herbivore resistance in plants should influence predator-herbivore interactions (19, 20), predator effects on herbivores were unrelated to milkweed resistance (fig. S3). In addition, the strength of predator effects on herbivores did not vary as a function of milkweed growth or growth response to soil fertility (table S2). However, both components of milkweed growth strategy convergently traded off against resistance so that species that were fast growing and responsive to soil fertility had low resistance (meaning, higher herbivore densities) (Fig. 2 and table S2), which is consistent with predictions from plant defense theory (17, 18). Thus, herbivore density was jointly and independently determined by means of interspecific variation in the top-down effect of predators and the bottomup effect of milkweed growth strategies.

Surprisingly, predator effects on plant biomass were unrelated to either predator effects on herbivores or herbivore density (resistance) (table S2). Instead, plant growth response to soil fertility predicted more than half of the variation in the top-down effects of predators on plant biomass (Fig. 2). At the same time, milkweed growth rate was not predictive of predator effects on plant biomass [and growth rate and response to soil fertility themselves are uncorrelated (table S2)]. Because the impact of predators on plant biomass was not related to the strength of herbivore suppression, variation in the indirect effects of predators on plants is probably attributable to variation in tolerance of milkweed species to herbivory. Consequently, an evolutionary trade-off leads to an association between high growth in response to soil fertility, low tolerance to herbivory, and an increase in predator effects on plant biomass.

Of several plant traits assayed, we found evidence suggestive of one mechanism behind the observed variation in the top-down effects of predators on plants. Plant emissions of sesquiterpene volatile organic compounds (VOCs) were significantly positively correlated with the top-down effects of predators on plant biomass (fig. S4 and table S3). Sesquiterpenes are a group of VOCs that can play a key role in indirect defense through recruitment of predators to plants (28). Therefore, variation among species in trophic cascade strength may be driven at least in part by interspecific variation in this ecologically important group of volatile compounds.

We have documented wide variation in topdown regulation of plant and herbivore biomass among a group of closely related species; this variation corresponded with a fundamental evo-





Fig. 2. (**A** to **D**) Relationship between milkweed species' growth rate, growth response to soil fertility, resistance to herbivores, and predator effects on plant biomass. For ease of interpretation, raw data are depicted with both raw and phylogenetically corrected R^2 values indicated in each panel. Where the phylogenetically independent correlations were significant [conducted by using generalized least-squares methods (27)], a linear best fit is shown through the raw data. Growth rate and response to soil fertility themselves are uncorrelated (table S2).

lutionary trade-off faced by plants. Interspecific variation in herbivore density was determined jointly and independently through variable effects of predators and two components of plant growth. In contrast, the cascading effects of predators on plant biomass were not tied to predator-herbivore interactions but instead fell along a probable trade-off between tolerance to herbivores and the bottom-up effects of soil fertility. Whereas food web models have predicted the correspondence of top-down and bottom-up effects on the basis of thermodynamic principles of energy flow (13, 14), we show here that such dynamics can similarly arise from convergent trade-offs faced by plants over evolutionary time. Our results underscore the importance of considering the plant-herbivore linkage as a determinant of trophic cascade strength, and the dynamic interplay between past evolutionary processes and contemporary ecological dynamics.

References and Notes

- O. J. Schmitz, P. A. Hambäck, A. P. Beckerman, *Am. Nat.* 155, 141 (2000).
- 2. F. Micheli, Science 285, 1396 (1999).
- M. T. Brett, C. R. Goldman, Proc. Natl. Acad. Sci. U.S.A. 93, 7723 (1996).
- 4. J. B. Shurin et al., Ecol. Lett. 5, 785 (2002).
- H. D. Vance-Chalcraft, J. A. Rosenheim, J. R. Vonesh, C. W. Osenberg, A. Sih, *Ecology* 88, 2689 (2007).

- A. Sih, G. Enlund, D. Wooster, *Trends Ecol. Evol.* 13, 350 (1998).
- 7. S. Nakano, M. Murakami, Proc. Natl. Acad. Sci. U.S.A. 98, 166 (2001).
- E. L. Preisser, D. I. Bolnick, M. F. Benard, *Ecology* 86, 501 (2005).

- 9. D. S. Gruner et al., Ecol. Lett. 11, 740 (2008).
- 10. M. Sipura, *Oecologia* **121**, 537 (1999).
- 11. R. E. Forkner, M. D. Hunter, *Ecology* **81**, 1588 (2000).
- I. Van der Stap, M. Vos, A. M. Verschoor, N. R. Helmsing, W. M. Mooij, *Ecology* 88, 2474 (2007).
- E. T. Borer, B. S. Halpern, E. W. Seabloom, *Ecology* 87, 2813 (2006).
- 14. L. Oksanen, S. D. Fretwell, J. Arruda, P. Niemela, *Am. Nat.* **118**, 240 (1981).
- J. T. Lill, R. J. Marquis, R. E. Ricklefs, *Nature* 417, 170 (2002).
- S. E. Helms, S. J. Connelly, M. D. Hunter, *Ecol. Entomol.* 29, 44 (2004).
- 17. P. V. A. Fine, I. Mesones, P. D. Coley, *Science* **305**, 663 (2004).
- P. D. Coley, J. P. Bryant, F. S. Chapin 3rd, Science 230, 895 (1985).
- 19. P. J. Ode, Annu. Rev. Entomol. 51, 163 (2006).
- 20. K. M. Clancy, P. W. Price, Ecology 68, 733 (1987).
- 21. J. P. Grime, Am. Nat. 111, 1169 (1977).
- J. Craine, *Resource Strategies of Wild Plants* (Princeton Univ. Press, Princeton, NJ, 2009).
- G. A. Polis, D. R. Strong, *Am. Nat.* **147**, 813 (1996).
 Materials and methods are available as supporting material on *Science* Online.
- 25. A. A. Agrawal, Am. Nat. 164, 113 (2004).
- L. V. Hedges, J. Gurevitch, P. S. Curtis, *Ecology* 80, 1150 (1999).
- 27. M. Pagel, *Nature* **401**, 877 (1999).
- 28. S. Rasmann et al., Nature 434, 732 (2005).
- A. A. Agrawal, M. Fishbein, Proc. Natl. Acad. Sci. U.S.A. 105, 10057 (2008).
- 30. We thank R. A. Smith and J. Goldstein for field assistance; S. Rasmann for contributing cardenolide data; M. Fishbein for providing phylogenetic information; and the University of California Berkeley Botanical Garden, S. Malcolm, A. Rapini, and M. Fishbein for providing plant material. The manuscript was improved by comments from A. Flecker, S. Rasmann, D. Raboski, and D. Gruner and supported by NSF, the Cornell Center for a Sustainable Future, and the University of California Irvine School of Biological Sciences.

Downloaded from www.sciencemag.org on March 25, 2010

Supporting Online Material

www.sciencemag.org/cgi/content/full/327/5973/1642/DC1 Materials and Methods Figs. S1 to S4 Tables S1 to S4

References

17 November 2009; accepted 16 February 2010 10.1126/science.1184814

A Peroxidase/Dual Oxidase System Modulates Midgut Epithelial Immunity in *Anopheles gambiae*

Sanjeev Kumar,*† Alvaro Molina-Cruz,* Lalita Gupta,† Janneth Rodrigues, Carolina Barillas-Mury‡

Extracellular matrices in diverse biological systems are cross-linked by dityrosine covalent bonds catalyzed by the peroxidase/oxidase system. We show that a peroxidase, secreted by the *Anopheles gambiae* midgut, and dual oxidase form a dityrosine network that decreases gut permeability to immune elicitors. This network protects the microbiota by preventing activation of epithelial immunity. It also provides a suitable environment for malaria parasites to develop within the midgut lumen without inducing nitric oxide synthase expression. Disruption of this barrier results in strong and effective pathogen-specific immune responses.

Insects, like most metazoa, harbor large numbers of commensal bacteria within their guts. Midgut epithelial cells need to protect the host from pathogenic organisms but must do so without mounting immune responses against the normal microbiota. This is instead they observed two. Although this result may not be exciting from a statistical point of view, it has been met with excitement in the field. For years, experiments have produced null results, providing only upper limits on cross sections of WIMP-nucleon interactions. The two detection events have raised hopes that more sensitive detectors may soon shed light on dark matter.

Because target materials can be made very pure, most radioactive backgrounds come from the surface of a detector, and instrumental effects are also stronger near detector edges. Continuing their two-decade long effort building improved detectors, the CDMS II Collaboration developed thicker germanium targets to improve the surfaceto-volume ratio, and with more efficient phonon collection to better reject surface events. This increases the acceptance of these new detectors to dark matter events and allows them to take the next step in sensitivity that is required to clarify the result reported.

Meanwhile, other experiments use differ-

ent technologies, each with its own difficulties and advantages. Some use liquid noble gases as target material and read out the scintillation light that is created in a particle interaction. If electrons liberated in the interaction drift through the liquid and are measured as well, this allows for background discrimination similar to CDMS. The major advantage of these liquid target detectors is that they can be made large, hence decreasing the surfaceto-volume ratio. The first of these to challenge the CDMS lead of the field was the XENON10 experiment (4). There, a large reduction of the radioactive background was achieved using only the innermost 5.4 kg from a 15-kg liquid xenon target. This year should see rapid progress, with at least three major liquid noble gas experiments expected to take and release data. Even ton-scale detectors will be feasible within a few years.

With all these highly sensitive detectors being or becoming operational, we can expect to learn soon whether or not CDMS observed dark matter and, with a bit of luck, what it is made of. Further, we expect that, should dark matter be WIMPs, we will detect many of them within only a few years and be able to estimate of their mass. Then the real fun will begin, as WIMP astronomy will allow us to infer otherwise inaccessible information. For example, due to their feeble interactions, the energy distribution of WIMPs in our galaxy, the Milky Way (see the figure), preserves features more than a billion years old that arose during collisions with other galaxies (5). Hence, measuring this WIMP energy distribution with direct dark matter detectors would open the history book of our galaxy.

References

- 1. E. Komatsu *et al., Astrophys. J. Suppl. Ser.* **180**, 330 (2009).
- The CDMS II Collaboration, *Science* **327**, 1619 (2010); published online 11 February 2010 (10.1126/science.1186112).
- 3. O. Adriani *et al.*, *Nature* **458**, 607 (2009).
- 4. J. Angle *et al.*, *Phys. Rev. Lett.* **100**, 021303 (2008).
- M. Vogelsberger et al., Mon. Not. R. Astron. Soc. 395, 797 (2009).

10.1126/science.1187972

ECOLOGY

A Green or a Prickly World?

Peter A. Hambäck

Is the world a green place where plant biomass abounds for herbivores to devour, or a prickly place where herbivores struggle to locate the few edible plant pieces? The answer to this question has crucial consequences for broad ecological and evolutionary questions. On page 1642 of this issue, Mooney *et al.* (1) show how evolutionary trade-offs among plant traits affect responses to herbivores and higher trophic levels.

If plant biomass is accessible but not consumed, herbivore numbers, and thus their consumption, may be regulated from the top by their natural enemies. If, in contrast, plant biomass is a nasty resource, then plant resistance to herbivory is central to understanding the greenness of the world. This dichotomy suggests a straightforward answer, but in nature, top-down forces interact in complicated ways with plant traits in determining the strength of species interactions. To address these issues, Mooney *et al.* have studied 16 related milkweed species. The results show that top-down effects from predator presence to plant biomass were determined not by plant resistance or by interaction strengths between herbivores and their natural enemies, but rather by a trade-off between plant growth strategy and resistance to herbivory.

The evolution of species traits depends on the main selective factors determining fitness. Populations facing high risks of predation will evolve different traits than populations where food limitation and starvation is common. In nature, this dependence is not unidirectional (even though most experiments are so), and traits themselves likely feed back to affect the species interactions and the fitness landscape. When we try to understand the role of certain species interactions in a specific ecological situation, what we see is a product of the evolutionary history of involved species and the proximate ecological factors. The interplay between these temporal scales for indirect interactions is only beginning to be disentangled by work such as that of Mooney et al.

In many insect-dominated ecological systems, plants emit volatile compounds to attract predators. These emissions create feedbacks between predators and herbivores that affect plant biomass and fitness (2). Predators and parasitoids attacking herbivorous insects use the chemical cues provided The evolution of plant traits shapes complex interactions with herbivores and their predators.

by plants at the time of herbivore attack to locate their prey. Mooney *et al.* (1) suggest that differences in volatile production—particularly of sesquiterpenes, which affect natural enemy attraction (3)—correlate with the effect of removing predators. Milkweed species that produced higher amounts of sesquiterpenes showed a larger difference in plant biomass with or without lady-bird beetles and other aphid predators, apparently because these species also differed in their tolerance to herbivory.

Most studies on the role of volatiles (the infochemical web) in plant-arthropod interactions have focused on mechanisms and chemical backgrounds, providing limited insights into the wider consequences for natural communities. Overlaying food webs with the infochemical web will increase our understanding of complex species interactions. In addition to the top-down effects on trophic cascades studied by Mooney et al., this was illustrated in a recent study of two related herbivorous beetles, their respective host plants, and a shared parasitoid (4). Field surveys suggested that mortality rates of one beetle species (Galerucella tenella) were higher when the other species (G. calmariensis) was pres-

Department of Botany, Stockholm University, SE-106 91 Stockholm, Sweden. E-mail: peter.hamback@botan.su.se

PERSPECTIVES



ent in the neighborhood. Seemingly, G. calmariensis was the better host for parasitoids, but adult parasitoids were nevertheless attracted A complex web. Food web interactions involve direct (solid lines) and indirect (hatched lines) effects. Predators and herbivores directly affect their resources through consumption, creating indirect effects of predators on plants (A). Predators also affect herbivore behavior and, indirectly, plant consumption (B) (*8*). Plants affect predator growth through effects on herbivore biomass and quality (C) (*9*) and predator search through damage-induced volatiles (D). Mooney *et al.* (1) suggest that trade-offs between plant growth strategy and resistance to herbivory moderate top-down effects (E).

to the scent of flowering meadowsweet (Filipendula ulmaria), host plant to G. tenella, and deposited more eggs in G. tenella larvae. As a result, attack was reduced and seed set increased on meadowsweet when growing together with the host plant of G. calmariensis, purple loosestrife (Lythrum salicaria). Herbivory from G. tenella can be very strong, defoliating the host plant, and selects for increases in defensive chemicals (5). This strong selection would presumably not occur in the presence of purple loosestrife.

Predators and parasitoids provide important ecosystem services to mankind and food production by reducing

pest densities. Estimates suggest that up to 15% of food production is lost to arthropod pests (6), and the global cost of biological

control is estimated at US\$417 billion/year (7). Yet, little is known about how to improve the conditions and efficiency of natural enemies in cropping systems. This development hinges on our detailed understanding of how complex interactions between plants, herbivores, and their predators determine the loss of plant tissue and the ability of plants to compensate for such losses. The integration of trophic dynamics and infochemical webs promises to provide deep additional insights to this end.

References

- 1. K. A. Mooney et al., Science 327, 1642 (2010).
- T. Bukovinszky, F. J. F. Van Veen, Y. Jongema, M. Dicke, Science 319, 804 (2008).
- M. R. Kant *et al., Adv. Bot. Res.* **51**, 613 (2009).
 J. A. Stenberg, J. Heijari, J. K. Holopainen, L. Ericson, *Oikos* **116**, 482 (2007).
- J. A. Stenberg, J. Witzell, L. Ericson, *Oecologia* 148, 414 (2006).
- 6. J. E. Losey, M. Vaughan, Bioscience 56, 311 (2006).
- 7. R. Costanza et al., Nature **387**, 253 (1997).
- 8. 0.]. Schmitz, Science **319**, 952 (2008).
- 9. P. J. Ode, Annu. Rev. Entomol. 51, 163 (2006).

Perovskite oxide catalysts may be able

10.1126/science.1187724

CHEMISTRY

Less Costly Catalysts for Controlling Engine Emissions

James E. Parks II

Devering the fuel consumption of transportation vehicles could decrease both emissions of greenhouse gases and our dependence on fossil fuels. One way to increase the fuel efficiency of internal combustion engines is to run them "lean," in the presence of more air than needed to burn all of the fuel. It may seem strange that engines are usually designed to run with fuel and air at stoichiometric balance, or even fuel rich. However, the way emissions have been controlled with catalytic converters has required some unburned fuel in the exhaust, especially for controlling the nitrogen oxide pollutants NO and NO₂ (called

 NO_x). On page 1624 of this issue, Kim *et al.* (1) report encouraging results for catalysts that can process NO_x in lean-burn engines. These perovskite oxide catalysts may help reduce or even eliminate the need for expensive and scarce platinum group metals (PGMs) in emission-control catalysts.

Commercially available vehicles with lean-burn technology have either diesel (2) or gasoline direct injection engines (3), and typically have additional costs of \$1000 to \$5000 compared with stoichiometric engine vehicles (4). A large part of the extra cost is due to engine modifications and exhaust catalysts that enable compliance with emission regulations.

Lean engines produce oxygen-rich exhaust, which prevents the reduction of NO₂

to control emissions from more fuel-efficient internal combustion engines and replace expensive noble metals.

via the "three-way" catalyst commonly used for stoichiometric engines. The new combustion and catalyst technologies for lean engines must meet NO_x emission standards as well as those for carbon monoxide, hydrocarbons, and particulate matter (soot). For diesel engines, special exhaust filters have eliminated the unsightly plumes of soot emissions from diesel exhaust.

Two catalyst technologies for controlling NO_x emissions from lean-burn engines are urea-based selective catalytic reduction (SCR) and the lean NO_x trap (LNT), also known as the NO_x storage and reduction catalyst. Urea SCR's main disadvantages are related to the requirement of onboard storage of the urea reductant and the need to heat the urea in cold weather. The issue of how

Fuels, Engines, and Emissions Research Center, Oak Ridge National Laboratory, Knoxville, TN 37932, USA. E-mail: parksjeii@ornl.gov



EDITED BY STELLA HURTLEY

Trophic Trade-Offs >>

There have been many attempts to document and explain the effects of predators on plant biomass in so-called "trophic cascades." Theory suggests that fast-growing plants are relatively undefended and suffer more from herbivory, which implies a functional trade-off between investment in traits relating to growth and defensive strategies. Mooney et al. (p. 1642; see the Perspective by Hambäck) compared responses to fertilization and aphid predators in 16 milkweed species. As predicted, interspecific variation in the strength of top-down control in terms of a tradeoff with growth was observed.



Platinum-Free Diesel

The efficiency advantages inherent in dieselbased combustion engines are counterbalanced by the production of pollutants such as nitrogen oxides (NO,). Currently, expensive precious metals, such as platinum, are required to remove these pollutants. Kim et al. (p. 1624; see the Perspective by Parks) show that a strontiumdoped perovskite catalyst, prepared from more abundant (and cheaper) elements, may help to lower the cost of NO_v treatments and thus ultimately make diesel a more cost-effective automotive fuel. Under conditions realistically simulating exhaust streams, the catalyst rivaled platinum in accelerating NO, decomposition.

Iron Exposure

The macrocyclic heme motif coordinates iron ions in proteins and plays a widespread role in biochemical oxidative catalysis. Bezzu et al. (p. 1627) prepared crystals in which analogous iron-centered macrocycles were aligned in pairs. The outer faces of the pairs exposed the iron ions to vacant cavities, where ligand exchange could take place; the inner faces were bound together by rigid bridging ligands lending the crystals structural integrity. The stability and high porosity of these crystals lend themselves to potential catalytic applications.

Preventing Radiation Damage

Inside a nuclear reactor, long-term exposure to radiation causes structural damage and limits the lifetimes of the reactor components. Bai et al. (p. 1631; see the Perspective by Ackland) now show, using three simulation methods able to cover a wide range of time and length scales,

that grain boundaries in copper can act as sinks for radiation-induced defects. The boundaries are able to store up defects, in the form of interstitials, which subsequently annihilate with vacancies in the bulk. This recombination mechanism has a lower energy barrier than the bulk equivalent, and so provides a lower-cost route for the copper to self-heal.

Fermion Behavior in an Optical Lattice

Due to their extreme tunability, optical lattices loaded with fermions and bosons are expected to

act as quantum simulators, answering complicated manybody physics questions beyond the reach of theory and computation. Some of these many-body states, such as the Mott insulator and the superfluid, have been achieved in bosonic optical lattices by simply changing the char-

acteristic depth of the lattice potential wells. Now, Hackermüller et al. (p. 1621) describe an unusual effect in an optical lattice loaded with fermions: When the strength of the attraction between the fermions is increased adiabatically, instead of contracting, the gas expands in order to preserve entropy.

Perfect Mismatch

Heteroepitaxy, or the overgrowth of one crystalline material onto a second crystalline material, is a key fabrication method for making thin films and nanoparticles. But if the lattice mismatch between the two materials is too large or messy, fractured interfaces result. Zhang et al. (p. 1634) describe a synthesis strategy to obtain spherical nanoparticles with a core-shell architecture that does not depend on heteroepitaxy. Silver was deposited and converted to various semiconductors through a series of chemical transformations to yield structurally perfect single-crystal semiconductor shells on a gold core, despite mismatches approaching 50%.

Lipid Kinase Revealed

The lipid kinase, Vps34, makes the key signaling lipid phosphatidylinositol 3-phosphate [PI(3)P] and has essential roles in autophagy, membrane trafficking, and cell signaling. It is a class III PI 3-kinase, a class against which there is currently no specific inhibitor. Miller

et al. (p. 1638) now describe the crystal structure of Vps34. Modeling substrate binding and combining structural data

of Vps34 with existing inhibitors might allow for the generation of inhibitors with high affinity and specificity.

Mosquito Double Act

binding and combining structural data with mutagenesis suggests a mechanism in which Vps34 is auto-inhibited in solu-tion, but adopts a catalytically active conformation on membranes. Structures f Vps34 with existing inhibitors might or the generation of inhibitors with high and specificity. **Quito Double Act** ase/dual oxidase (duox) systems act in to catalyze the nonspecific formation of ne bonds, which cross-link a variety of s. Knowing that these reactions are d in fine-tuning insect immune ses, **Kumar et al.** (p. 1644, published 11 March) investigated how the peroxidase/ stem in malaria-vector mosquitoes pro-Peroxidase/dual oxidase (duox) systems act in concert to catalyze the nonspecific formation of dityrosine bonds, which cross-link a variety of proteins. Knowing that these reactions are involved in fine-tuning insect immune responses, Kumar et al. (p. 1644, published online 11 March) investigated how the peroxidase/ duox system in malaria-vector mosquitoes pro-

26 MARCH 2010 VOL 327 SCIENCE www.sciencemag.org Published by AAAS

1 SUPPLEMENTARY MATERIALS

2 <u>Methods</u>

3 Six-week old plants of 16 closely related milkweed species (S1) were out-planted 4 to a plowed field near Ithaca, NY USA on 7 June 2007. Fourteen species were in a two-5 by-two factorial experimental design of soil fertilization and predator manipulation, while 6 A. candida and A. nivea were only in the latter treatment due to limited sample size. The 7 native range of these milkweeds spans North to South America, including a variety of 8 open habitats. Seeds came variously from commercial seed sources and field collections. 9 We obtained seedlings of A. mexicana adjacent to mature plants in the collection of the 10 U.C. Berkeley Botanical Garden and A. barjonifolia was propagated from cuttings of 11 mature plants maintained in a greenhouse. 12 The aphid predators observed in this experiment were dominated by larval and 13 adult coccinellids (Coccinellidae: Coleoptera) and larval syrphids (Syrphidae: Diptera), 14 both of which are generalists that feed upon aphids world-wide (S2). There were few 15 visible signs of aphid parasitism. Aphis nerii is notable for the fact that it reproduces 16 exclusively via parthenogenesis and is thus genetically uniform (S3). Consequently, the 17 arthropod community associating with milkweeds at the field site in Ithaca is similar to 18 that occurring in the native ranges of the studied species. 19 In the field, fertilized plants received 5.0 g of slow-release 14-14-14 NPK 20 fertilizer. Predator density was reduced by 77% using buried spun polyester bags, with 21 control plants receiving identical bags with vertical slits to allow predator access. There

22 were five plants of each milkweed species in each experimental treatment, and

assignments of all treatments to positions within the experimental plot were random.

Colonies of 12 *Aphis nerii* aphids were initiated on 19 June. Aphid analyses are based
upon counts made on 13 July, during peak aphid density, and above-ground plant
biomass was harvested on 1 August. Relatively few aphids were of winged dispersal
morphs, suggesting that aphid movement among plants was relatively unimportant.
Milkweed herbivores other than *A. nerii* were rare, with individuals being observed in
only 3% of plant surveys.

30 Data on species cardenolide concentration, trichome density, and specific leaf 31 area were taken from other experiments on the same milkweed species (S4, S5). Volatile 32 emissions were measured on 2-3 July from 3 to 5 randomly selected individuals of each 33 species in treatments exposed to predators and without fertilizer. Aphids infested all 34 plants, but damage by other herbivores was minimal. Volatiles were collected on 35 activated charcoal adsorption tubes (Supelco, Bellefonte, PA, USA), eluted with 36 dichloromethane and analyzed by gas chromatography-mass spectrometry as described 37 elsewhere (S6). Individual compounds were classified into compound classes of 38 monoterpenes (ocimene, myrcene and linalool), green leaf volatiles (Z3-hexenal and Z3-39 hexenyl acetate) and sesquiterpenes including trans- β -caryophyllene, farnesene, nerolidol 40 and 5 unidentified sesquiterpenes based on their mass spectra.

41 Tests were performed for effects of milkweed species, fertilization, predator 42 exclusion and their two- and three-way interactions on aphid density (transformation for 43 normality of residuals = ln([aphid count+1]/plant dry biomass) and plant biomass 44 (transformation for normality of residuals = plant dry biomass^0.5) by ANOVA. Results 45 for plant biomass were qualitatively identical when based upon a log transformation.

46	Where ANOVA showed species differences, raw and phylogenetically corrected				
47	correlations were performed among log response ratio (LRR) effect sizes and species				
48	traits (S7). Effect sizes for predator effects were calculated across both soil fertility				
49	treatments, as were soil fertility effects calculated across both predation treatments. In				
50	some cases (those shown in Table S2) LRRs and species means for herbivore resistance				
51	and growth were non-independent due to their calculations being based on some of the				
52	same data (e.g., plant biomass was a component of both aphid density and plant growth				
53	rate). Consequently, the significance of these raw correlations was tested by				
54	randomization (S8). In these randomization tests, the species means for plant biomass				
55	and aphid abundance from each level of the predator exclusion and soil fertility				
56	treatments were randomized across species, after which all dependent variables and				
57	correlation coefficients were recalculated. The observed correlation coefficients were				
58	then compared to the distribution of 999 correlations generated through this				
59	randomization process.				
60	Phylogenetic analyses where only performed where randomization tests showed				
61	the raw correlations to be significant (P<0.05) and unbiased (i.e. a mean randomized				
62	correlation coefficient not differing significantly from 0) because there are no developed				
63	methods for controlling for such bias in phylogenetic independent contrasts. Where the				
64	pairs of correlated variables were independent (those shown in Table S3), phylogenetic				
65	analyses were performed for all correlations. Phylogenetic analyses were conducted				
66	with the Continuous module of BayesTraits (<u>http://www.evolution.rdg.ac.uk/</u>				
67	BayesTraits.html) (S9) and the generalized least squares phylogenetically corrected				
68	correlations were parameterized to equal the traditional independent contrast.				

69	Correlations were performed among LRR of predator effects on aphid density and				
70	plant biomass, LRR of soil fertility effects on plant biomass, milkweed species mass and				
71	herbivore resistance (quantified as -1 x aphid density in the absence of predators). Among				
72	these 10 correlations, five were significant, which is statistically improbable by chance				
73	alone (P<0.0001, binomial expansion test). Similar analyses were performed between				
74	species traits and LRR of predator effects on aphid density and plant biomass, LRR of				
75	soil fertility effects on plant biomass and herbivore resistance. Among these 28				
76	correlations five were significant, which is statistically improbable by chance alone				
77	(P=0.009, binomial expansion test).				
78					
79	Tables				
80	Table S1. ANOVA tables for results of statistical tests for milkweed species,				
81	predator and fertilization effects on herbivore density and milkweed growth response.				
82	Table S2. Correlations among growth, resistance and milkweed response to				
83	manipulation of predators and soil fertility.				
84	Table S3. Phylogenetically corrected correlations of milkweed species traits with				
85	herbivore performance and predator and soil fertility effects.				
86					
87	Figures				
88	Figure S1. Mean \pm 1SE herbivore density (bars, left Y axis,) and herbivore				
89	abundance (circles, right Y axis) for each of 16 milkweed species. Herbivore means are				
90	across all levels of soil fertility and predator manipulations. Milkweed species are				

91 arranged in order of increasing size, with mean species g dry biomass listed in

92 parentheses.

- 93 Figure S2. Interactive effects of predators and soil fertility on milkweed biomass.
 94 Mean ± 1SE plant biomass in each treatment.
- Figure S3. Relationship between milkweed species' resistance to herbivores and
 the strength of predator effects on those herbivores. For ease of interpretation, raw data
 are depicted and raw and phylogenetically corrected R² are indicated.
- 98 **Figure S4.** Relationship between milkweed species' production of sesquiterpene
- 99 volatiles and the strength of predator effects on plant biomass. For ease of interpretation,
- 100 raw data are depicted and raw and phylogenetically corrected R^2 are indicated.
- 101

102 <u>References</u>

- 103 S1. A. A. Agrawal, M. Fishbein, Proc. Natl. Acad. Sci. U. S. A. 105, 10057 (2008).
- 104 S2. A. F. G. Dixon, Aphid ecology : an optimization approach. (Chapman & Hall,
- 105 London, ed. 2nd, 1998), pp. viii, 300.
- 106 S3. R. L. Blackman, V. F. Eastop, Aphids on the world's herbaceous plants and shrubs.
- 107 (Wiley, London, 2006), vol. 2, pp. 1439.
- 108 S4. A. A. Agrawal et al., New Phytol. 183, 848 (2009).
- 109 S5. A. A. Agrawal et al., PNAS 106, 18067 (2009)
- 110 S6. A. Kessler, I. T. Baldwin, *Science* **291**, 2141 (Mar 16, 2001).
- 111 S7. L. V. Hedges, J. Gurevitch, P. S. Curtis, *Ecology* **80**, 1150 (Jun, 1999).
- 112 S8. M. T. Brett, Oikos 105, 647 (2004).
- 113 S9. M. Pagel, *Nature* **401**, 877 (Oct, 1999).

	Source	Degrees of freedom	Sums of squares	Mean Square	F Value	Р
Aphid density	Model	59	261.93	4.44	3.5	<.0001
	Error	251	318.57	1.27		
	Corrected Total	310	580.50			
	Pred	1	41.93	41.93	33.04	<.0001
	Fert	1	10.68	10.68	8.41	0.0041
	Pred*Fert	1	0.05	0.05	0.04	0.8412
	Species	15	151.65	10.11	7.97	<.0001
	Pred*Species	15	35.75	2.38	1.88	0.0258
	Fert*Species	13	5.22	0.40	0.32	0.9892
	Pred*Fert*Species	13	14.68	1.13	0.89	0.5647
Plant	Model	59	55530.28	941.19	14.61	<.0001
biomass	Error	245	15787.21	64.44		
	Corrected Total	304	71317.49			
	Pred	1	1691.86	1691.86	26.26	<.0001
	Fert	1	2083.96	2083.96	32.34	<.0001
	Pred*Fert	1	311.12	311.12	4.83	0.0289
	Species	15	44161.50	2944.10	45.69	<.0001
	Pred*Species	15	2485.98	165.73	2.57	0.0014
	Fert*Species	13	2643.06	203.31	3.16	0.0002
	Pred*Fert*Species	13	698.22	53.71	0.83	0.6243

Table S1. ANOVA tables for results of statistical tests for milkweed species, predator and fertilization effects on herbivore density and plant biomass.

	growth rate	herbivore resistance	predator LRR on herbivores	predator LRR on plant biomass	fertilization LRR on plant biomass
		-0.81**			
growth rate					
herbivore resistance	-0.44*				-0.79**
predator LRR on herbivores	0.20	0.49			
predator LRR on plant biomass	0.02	-0.55	-0.35		0.75**
fertilization LRR on plant biomass	0.30	-0.48*	-0.22	0.64**	

Table S2. Correlations among growth, resistance and milkweed response to manipulation of predators and soil fertility.

1. Phylogenetically corrected correlation coefficients (above diagonal) are shown when raw correlation coefficients (below diagonal) are statistically significant in randomization tests. Randomization tests were used because variables are non-independent due to their being based on some of the same data (See Methods in the Supplement for details).

2. Sample size for correlations with fertilization LRR on plant biomass are N=14, all others are N=16.

3. * P<0.05; ** P<0.01; ***P<0.001

4. Correlations with fertilization effects on herbivores are not shown because these effects did not vary significantly among milkweed species (Table S1).

	Species effect	Predato	Fertilization effect	
	aphid	aphid	plant	plant
Species trait	density	density	biomass	biomass
Plant dry biomass	0.65*	0.33	0.24	0.63*
Cardenolide equivalents	-0.05	0.27	-0.06	0.07
Specific Leaf Area	0.04	-0.09	0.28	-0.52*
Trichome density	-0.25	0.24	-0.12	-0.03
Volatile monoterpenes	0.08	0.11	0.04	0.13
Green leaf volatiles	0.23	-0.14	0.36	0.39
Volatile sesquiterpenes	0.25	-0.03	0.58*	0.44(*)
1 (*) D 0 0 7 1 D (10 + D = 0.07			

Table S3. Phylogenetically corrected correlations of milkweed species traits with herbvore performance and predator and soil fertility effects.

1. (*) P>0.05 and P<0.10; * P<0.05



Milkweed species in order of size (dry mass g)



Figure S3



Figure S4

